



What does positron emission tomography offer oncology?

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Abstract

The origins of positron emission tomography (PET) date back 70 years. Since the 1970s, however, its use has increased exponentially in the fields of neurology, cardiology and oncology. [^{18}F]-Fluorodeoxyglucose (FDG) whole-body scanning is by far the most widely utilised and recognised application of PET in oncology. However, PET is a very versatile and powerful imaging modality capable of helping bridge the gap between the laboratory and the clinic. This article reviews the history and current applications of PET in oncology and then explores some of the newer applications and potential future uses of this versatile technology particularly in the area of cancer research. © 2000 Elsevier Science Ltd. All rights reserved.

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1. Positron emission tomography (PET)

Positron emission tomography (PET) is a sophisticated imaging technique that is able to detect, localise and quantitate radionuclides in the body non-invasively. PET utilises commonly radionuclides such as [^{15}O], [^{11}C], [^{18}F], [^{124}I] and incorporates them into molecules in a potentially huge range of compounds ranging from simple H_2O to complex pharmaceuticals. These compounds can then be used to image important physiological, biochemical and molecular processes at a level of sensitivity unmatched by other imaging techniques (see Fig. 1) [1].

2. History of PET in oncology

The beginning of the development of PET dates back to early this century when the theoretical physicist P. Dirac postulated the existence of positive electrons based on the equations of quantum mechanics and Einstein's theory of relativity [2]. C.D. Anderson subsequently proved Dirac's theory in 1932 when he observed experimentally that cosmic rays include particles with

the mass of electrons, but with a positive charge [3]. These particles were called positrons. Around this time in Berkeley, California Ernest Lawrence and his team were developing the first cyclotron: two D-shaped magnets capable of accelerating particles to produce progressively higher energy protons and deuterons that could then bombard elements to explore the nature of the atomic nucleus [4]. The fusion of the theory and experimental observation led to the development of larger cyclotrons able to produce large quantities of artificial radioisotopes such as carbon-11, nitrogen-13, oxygen-15 and fluorine-18, isotopes that continue to form the backbone of medical PET imaging today.

Another step in the evolution of the PET scanner we know today was the development in the 1940s of a system for external measurement of radiotracers. Hand-held Geiger–Muller counters were initially used to measure the rate of accumulation of radioactive iodine by the thyroid gland to help decide whether a thyroid nodule was benign or malignant [4]. An automated system of detector movement followed producing the first so-called 'scanner' [5]. It was not long before this was used to produce nuclear images of other organs and *in vivo* molecular imaging was born [4]. Multidetector systems were then developed culminating in the ring detector systems still used today.

The next major step in PET development for oncological use occurred in the 1970s. In 1948, Kety and

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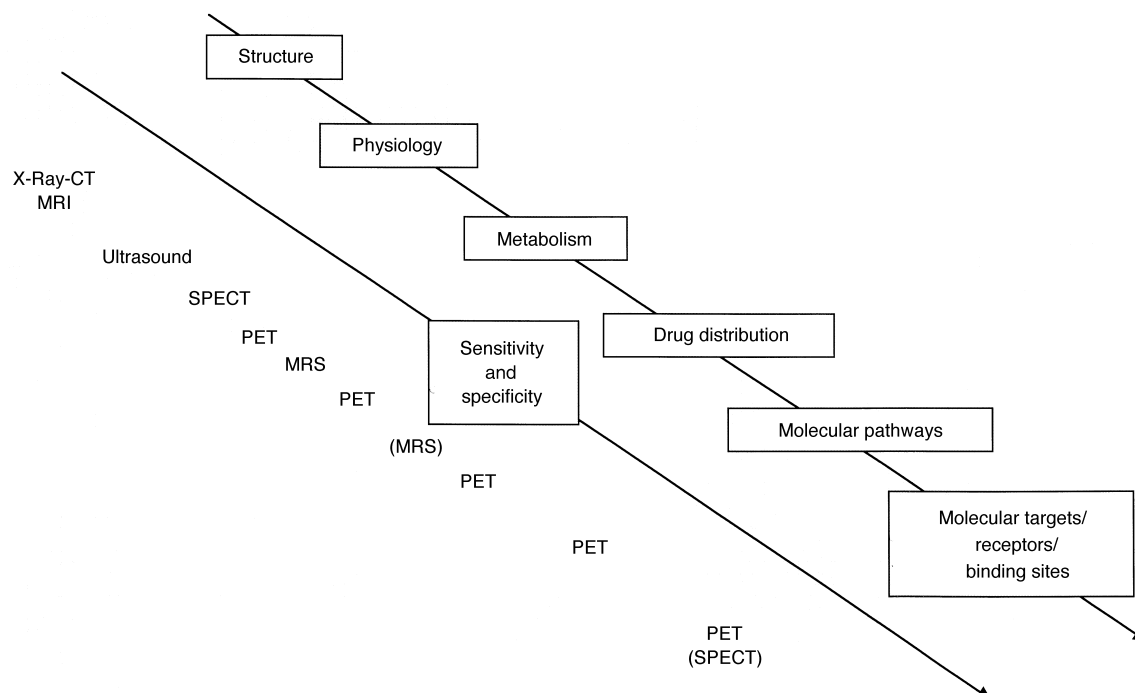


Fig. 1. The spectrum of medical imaging, which ranks the ability to image decreasing concentrations (sensitivity) of specific molecules (specificity). The spectrum covers a sensitivity range of 10^9 (millimolar to picomolar). CT, computed tomography; MRI, magnetic resonance imaging; MRS, magnetic resonance spectroscopy; PET, positron emission tomography; SPECT, single photon emission computed tomography. Reproduced with permission [1].

Schmidt had used nitrous oxide to apply the Fick principle to measure cerebral blood flow and oxygen utilisation [6]. This method of measuring cerebral blood flow made it possible to determine in living persons the average rate of glucose use in the brain as a whole from measurements of blood flow and the arteriovenous difference in glucose levels and oxygen utilisation. In 1977, Sokoloff described the carbon-14 deoxyglucose method for measurement of local cerebral glucose utilisation [7]. This technique was subsequently adapted to use fluorine-18 deoxyglucose [8] and the [^{18}F]-fluorodeoxyglucose (FDG) scan for the measurement of glucose metabolism, *in vivo*, in humans was born.

3. Development and current status of [^{18}F]-FDG imaging

The use of FDG in PET is based on the Warburg's observation back in the 1930s that malignant tumours have an increased glycolytic rate [9]. FDG initially follows the same metabolic pathway as glucose. It is carried into the cell by endothelial glucose transports and is converted to FDG-6-phosphate. Unlike glucose, this is then trapped in the cell, where it accumulates at a rate proportional to glucose utilisation [10]. In the 1980s, a number of animal studies demonstrated an increased uptake of FDG in transplanted and spontaneous tumours [11–13]. Di Chiro and colleagues [14] was one of the first to show that FDG–PET could differentiate

tumour recurrence from post-radiotherapy changes in the brain. With the development of the PET whole-body scanner, the range of applications of FDG expanded and its use is becoming more widespread in a number of clinical applications (Table 1). Some of these uses have been more extensively investigated and evaluated than others and there are many ongoing studies, not only in the clinical applications, but also in the refinement of the technique itself.

4. Future developments in the field of FDG–PET

One of the main criticisms of the clinical use of FDG–PET has been the lack of a standardised methodology in the data collection and analysis of scans. For example, there are several methods of evaluating whole-body FDG–PET scans: subjective, qualitative visual evaluation, a more time-consuming and costly method of quantitating the standard uptake value or SUV with Patlak analysis, a semi-quantitative method using an SUV curve, other kinetic parameters of glucose uptake such as K_i , the net influx constant, and M_{rglu} , the glucose metabolic rate. There is variation in the timing of scanning protocols and whether lean body mass or total body mass is used in the calculations of SUVs. Attempts are being made to standardise the methodology [46] and such efforts are essential in improving the quality and reliability of FDG–PET in the future.

An aspect of FDG–PET that is being increasingly studied is the use of this technique to assess response to treatment [10,47]. This has potential advantages for patient management and drug development. A paper by Brock and associates [38] performed FDG–PET measurements in brain tumours at 7 days after treatment and found that these results could predict ultimate clinical and radiological response recorded at 2 months. Another example is in the paralleling of phase I studies with [¹⁸F]-FDG–PET, which may then be used to predict toxicity and tumour response at a preclinical level. This can then be used to facilitate effective drug development and in defining optimal drug scheduling and in the individualisation of therapy [10]. Recent European guidelines for the use of [¹⁸F]-FDG for response assessment in oncology have been produced by the European Organization for Research and Treatment of Cancer (EORTC) PET study group [46].

[¹⁸F]-FDG–PET continues to play an increasing part in the diagnosis and management of patients with cancer. In the USA for example, in lung cancer, [¹⁸F]-FDG has become the standard investigation in the diagnosis of solitary lung nodules and in the pre-operative staging of lung carcinoma. Validation in other areas (see Table 1) will continue and it may become increasingly incorporated into patient management algorithms. Its relative place in diagnostics has been discussed elsewhere [48].

[¹⁸F]-FDG has dominated the PET oncology field, but perhaps the greatest strength in PET is the ability to utilise many other radioisotopes; isotopes that can be

incorporated into complex compounds and pharmaceuticals and used to investigate the biology of human cancers *in vivo* and the effects of various manipulations.

4.1. Tumour physiology

Measuring blood flow, blood volume and oxygen utilisation using ¹⁵O-labelled H₂¹⁵O and C¹⁵O has been a mainstay of PET studies from the early years. Initially these were developed for brain studies [49,50] but the technique has subsequently been modified to measure blood flow and exchanging water space in breast tumours [51]. These techniques have a potentially important role in the development and assessment of new antivasular and antiangiogenic therapies targeting tumour vasculature that are being increasingly investigated.

5. Imaging of tumour metabolism

Tumours are characterised by abnormal growth and metabolism. However, [¹⁸F]-FDG imaging is not completely specific for malignant tumours: inflammation, tuberculosis and certain non-malignant tumours have increased uptake. In an attempt to increase the specificity of PET in imaging cellular growth, other radio-tracers were developed and are continuing to be developed. [¹¹C]-thymidine has been developed as a specific marker of DNA synthesis. Data from normal tissue models have shown correlation between [¹¹C]-thymidine uptake as determined by PET, DNA turnover in regenerating tissues and non-regenerating livers and cell proliferation [52,53]. Preliminary data suggest that [¹¹C]-thymidine uptake may provide a more direct and relevant *in vivo* measure of response in both tumours and normal tissue [54,55]. However, the rapid metabolism and accumulation of metabolites complicates the interpretation of [¹¹C]-thymidine scans. New tracers are being developed that enter the DNA synthetic pathway but are more stable to systemic degradation e.g. [¹⁸F]-3'-deoxy-3'-fluorothymidine (FLT) [56] and bromine-76-bromodeoxyuridine [57].

Tracers of protein synthesis are also being developed as markers of biosynthesis [58]. [Methyl-¹¹C]-methionine] is the most widely used amino acid radio-pharmaceutical [59,60]. However, again their secondary metabolism [61] and dilution into unknown pool sizes of intracellular amino acids [58] makes their interpretation difficult.

A specific, easy to use marker of tumour growth would have an important role in the oncological use of PET [54]: It could be used to accurately assess early response and predict clinical outcome of treatment reducing the exposure to patients and the expense of ineffective treatments. It could be used to confirm com-

Table 1
A list of clinical applications where there is good evidence that [¹⁸F]-FDG–PET improves the management of certain tumours

Clinical applications of FDG–PET	Tumours with evidence of benefit [Ref.]
Preoperative detection and staging of disease	Lung cancer [15–17] Breast cancer [18,19] Colorectal cancer [21,22] Melanoma [22,23] Head and neck cancer [24,25] Pancreatic cancer [26]
Detection of recurrence	Lymphoma [27] Head and neck cancer [28] Thyroid cancer [29,30]
Differentiation between recurrence and scarring post-treatment	Colorectal cancer [31,32] Brain tumours [33,34]
Evaluating response to treatment	Head and neck cancer [35–37] Brain tumours [38] Sarcoma [1,39] Breast cancer [40,41]
Determination of biopsy site	Head and neck cancer [42,43] Brain tumours [44,45]

FDG–PET, [¹⁸F]-fluorodeoxyglucose positron emission tomography.

plete response to therapy. It could be used when carrying out trials of new treatments, schedules or combinations providing a quantitative measure of fractional cell kill. And it could be useful in assessing the *in vivo* response of normal tissue to therapy.

6. Tumour receptor imaging

The last 20 years have seen an exponential increase in our knowledge and understanding of tumour cell biology and molecular characteristics of tumours and tumorigenesis. Amongst this has been recognition that receptor and transport systems play an important part in tumour biology and are a potential target for future therapies. Radiopharmaceuticals that can assess such markers are being developed to aid diagnosis and therapeutic planning. Several fluorinated antioestrogens and anti-androgens have been labelled as PET-tracers and measurement of receptor concentrations made [62,63]. Oestrogen receptor (ER) status of breast tumours has prognostic implications and currently the method of assessing ER status is from biopsy specimens. To date, the use of PET constitutes the most reliable non-invasive method for assessing ER status [64]. An interesting development along this whole line uses ^{18}F -labelled and ^{131}I -labelled tamoxifen analogues to image ER-positive breast tumours [65]. The potential use of such ligands is to predict a patient's susceptibility to tamoxifen, determine the proportion of oestrogen receptors occupied by therapeutic drug and monitor the effectiveness of hormonal therapy on an ongoing basis [64]. Similar uses have been suggested for androgen receptor ligands in prostate cancer [66]. Potential targets in the future include other receptors that are becoming recognised as being important in oncology such as the vascular endothelial growth factor (VEGF), epidermal growth factor (EGF), P-glycoprotein (Pgp), a plasma membrane transporter and other protein products encoded by multidrug resistant (MDR) genes are another potentially important area of PET research. These proteins reduce intracellular accumulation of cytotoxic agents and the efficacy of chemotherapy. A number of PET radiopharmaceuticals are being developed to investigate the process further and in conjunction with strategies to block expression or activity of these proteins to improve the efficacy of cytotoxic agents. These radiopharmaceuticals include Technetium-99m sestamibi which is a substrate for Pgp and has been used in clinical studies for tumour imaging, and to visualise blockade of Pgp-mediated transport after modulation of the Pgp pump [67]. ^{11}C -labelled drugs such as ^{11}C -verapamil and ^{11}C -daunorubicin [68] and ^{11}C -colchicine [69] have also been investigated and are additional tools for the quantification of Pgp-mediated transport with PET *in vivo*. A potential use of such

agents would be to select patients who may benefit from the addition of Pgp modulators, which are being developed [70].

7. Tumour hypoxia

Hypoxia is present in almost all tumours *in vivo* [71]. It is one of the main causes of resistance to conventional radiotherapy and chemotherapy and hence an important factor to consider when investigating tumours and developing treatments. Tracers such as ^{18}F -fluoromisonidazole already exist that can quantitate hypoxia in tumours using PET [71–73]. Newer, more sensitive agents are being developed [74] and have a potential role in the selection of patients for therapy with bio-reductive agents, antiangiogenic agents, antivascular agents and hypoxia-targeted gene therapy [75].

8. Imaging gene expression

Gene therapy has been hailed as one of the next major developments in cancer therapeutics. It has not, as yet, translated into clinical application, but hopes are still high. PET could have a unique role to play in development and assessment. Most work to date has centred on gene therapy in tumours transduced with the herpes simplex virus type 1 thymidine kinase (*HSV1-tk*) suicide gene. The principle of suicide genes is that they mediate conversion of a prodrug only on reaching the site of its intended target [76]. In the case of *HSV1-tk* the prodrug is gancyclovir. Cells that express *HSV1-tk* are more susceptible to gancyclovir toxicity because they phosphorylate the drug, which then becomes trapped, additionally phosphorylated and incorporated into cellular DNA, leading to chain termination [64]. A number of analogues to gancyclovir that can be imaged by PET have been developed [77–79] and used to monitor gene expression in animals. In future, these will be developed for use in humans.

9. Drug development

A number of conventional cytotoxic agents have already been radiolabelled and have proved useful in investigating aspects of drug pharmacokinetics that cannot be measured in any other way. Conventional pharmacokinetic studies rely on inferring information on tissue levels from plasma measures. PET, however, can be used to directly measure the drug concentration in tissue. For example, ^{57}Co -bleomycin has been used to produce tissue and plasma time-activity curves that have been used to determine tumour and normal tissue kinetics directly [80]. The distribution of ^{13}N -cisplatin

[81], 1,3-bis(2-chloroethyl)-1-nitrosourea (BCNU) and 2-chloroethyl-3-sarcosinamide-1-nitrosourea (SarCNU) [82] in brain tumours has been investigated. More complicated pharmacokinetic analyses have been performed with ^{18}F -labelled 5-fluorouracil. Not only has information been obtained on the pharmacokinetics of the drug in metastases and normal tissue [83], but this has been applied to predicting response to therapy [84,85]. ^{18}F -5-FU can also be used to assess the effects of biochemical modulation of 5-FU [75,86] *in vivo* (see Fig. 2) [87].

There is unlimited potential for the investigation of drugs in oncology using such methods. What is needed and what will be developed are improvements in the whole radiolabelling process: basic radiochemistry, precursor synthesis, rapid radiolabelling, automation of radiosynthesis and analytical techniques [70].

10. Radioimmunotargeting and antibody imaging

Therapeutic monoclonal antibodies are establishing themselves in clinical practice and are having some success [88,89] particularly in the area of lymphoma and breast cancer. The high level of interest in such agents has led to recent advances in radioimmunotargeting. This involves the administration of nuclide-carrying antibody with a specific target that administers a low

dose of radioactivity over a long period of time. Most radionuclides used in radioimmunotherapy have positron-emitting analogues, which can be used for PET imaging [90] and for performing dosimetry. With the expansion of this area of cancer therapeutics, PET will have an important role.

11. Summary

The remit of this article was to introduce the reader to some of the accomplishments of PET in the area of oncology and to look ahead to its future. Space does not allow a comprehensive review, but we have attempted to present a broad illustration of the many and varied uses of PET in the oncology field. Although FDG–PET is an important area, it does tend to dominate peoples' perception of PET. FDG–PET will continue to be used as a diagnostic tool and as an adjunct to the other imaging modalities. The real strength of PET, however, lies in its unique ability to quantitatively assess biochemical and physiological processes *in vivo*. The future of oncology lies in individualisation of therapy; being able to select the best treatment for an individual patient; being able to objectively assess response at an early stage; being able to assess normal tissue toxicities. It lies in more specific tumour targeting and improvements in delivery and scheduling of anticancer therapies. PET is a

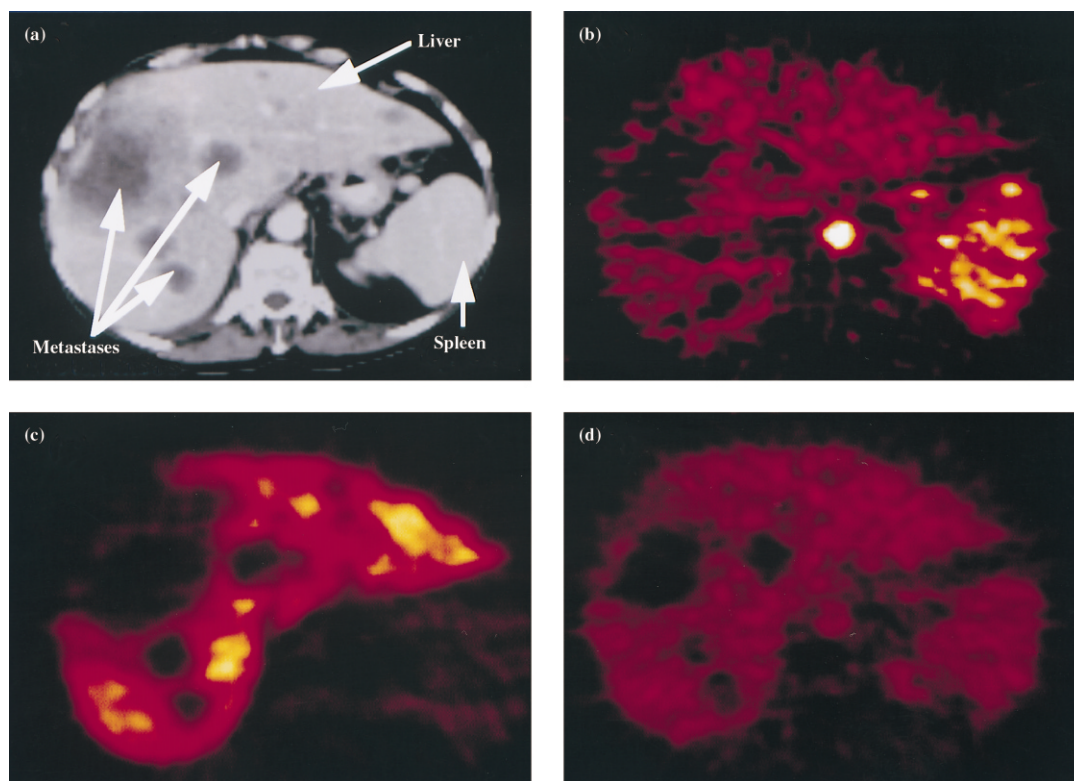


Fig. 2. Transabdominal computed tomography (a) and corresponding positron emission tomography (PET) blood flow (b) and PET ^{18}F -fluorouracil images without eniluracil (c) and after eniluracil (d), showing liver, spleen and multiple hepatic metastases. Reproduced with permission [87].

valuable tool in all these areas and the future for PET in oncology, above all, looks healthy.

References

- Jones T. The imaging science of positron emission tomography. *Eur J Nucl Med* 1996, **23**, 807–813.
- Dirac PAM. A theory of electrons and proton. *Proc Roy Soc* 1930, **A126**, 360–365.
- Anderson CD. Energies of cosmic-ray particles. *Phys Rev* 1932, **40**, 405–421.
- Wagner HNJ. A brief history of positron emission tomography (PET). *Semin Nucl Med* 1998, **28**, 213–220.
- Cassen B, Curtis L, Reed CW. A sensitive directional gamma-ray detector. *Nucleonics* 1950, 78–81.
- Kety SS, Schmidt CF. The nitrous oxide method for the quantitative determination of cerebral blood flow in man theory, procedure and normal values. *J Clin Invest* 1948, **1**, 476–483.
- Sokoloff L, Reivich M, Kennedy C, et al. The [^{14}C]deoxyglucose method for the measurement of local cerebral glucose utilization: theory, procedure, and normal values in the conscious and anesthetized albino rat. *J Neurochem* 1977, **28**, 897–916.
- Reivich M, Kuhl D, Wolf A, et al. Measurement of local cerebral glucose metabolism in man with ^{18}F -2-fluoro-2-deoxy-D-glucose. *Acta Neurol Scand Suppl* 1977, **64**, 190–191.
- Warburg O. The metabolism of tumors. In Smith R, ed. New York, Richard R. Smith Inc., 1931, 129–169.
- Brock CS, Meikle SR, Price P. Does fluorine-18 fluorodeoxyglucose metabolic imaging of tumours benefit oncology? *Eur J Nucl Med* 1997, **24**, 691–705.
- Som P, Atkins HL, Bandyopadhyay D, et al. A fluorinated glucose analog, 2-fluoro-2-deoxy-D-glucose (F-18): nontoxic tracer for rapid tumor detection. *J Nucl Med* 1980, **21**, 670–675.
- Fukuda H, Matsuzawa T, Abe Y, et al. Experimental study for cancer diagnosis with positron-labeled fluorinated glucose analogs: [^{18}F]-2-fluoro-2-deoxy-D-mannose a new tracer for cancer detection. *Eur J Nucl Med* 1982, **7**, 294–297.
- Abe Y, Matsuzawa T, Fujiwara T, et al. Assessment of radiotherapeutic effects on experimental tumors using ^{18}F -2-fluoro-2-deoxy-D-glucose. *Eur J Nucl Med* 1986, **12**, 325–328.
- Di Chiro G, DeLaPaz RL, Brooks RA, et al. Glucose utilization of cerebral gliomas measured by [^{18}F] fluorodeoxyglucose and positron emission tomography. *Neurology* 1982, **32**, 1323–1329.
- Weng E, Tran L, Rege S, et al. Accuracy and clinical impact of mediastinal lymph node staging with FDG–PET imaging in potentially resectable lung cancer. *Am J Clin Oncol* 2000, **23**, 47–52.
- Berlangieri SU, Scott AM, Knight SR, et al. F-18 fluorodeoxyglucose positron emission tomography in the non-invasive staging of non-small cell lung cancer. *Eur J Cardiothorac Surg* 1999, **16**(Suppl. 1), S25.
- Marom EM, McAdams HP, Erasmus JJ, et al. Staging non-small cell lung cancer with whole-body PET. *Radiology* 1999, **212**, 803–809.
- Hoh CK, Schiepers C. ^{18}F -FDG imaging in breast cancer. *Semin Nucl Med* 1999, **29**, 49–56.
- Smith IC, Ogston KN, Whitford P, et al. Staging of the axilla in breast cancer: accurate in vivo assessment using positron emission tomography with 2-(fluorine-18)-fluoro-2-deoxy-D-glucose. *Ann Surg* 1998, **228**, 220–227.
- Ott DJ. Whole-body PET proves superior to CT for staging primary colorectal carcinoma. *Am J Gastroenterol* 1999, **94**, 285–286.
- Abdel-Nabi H, Doerr RJ, Lamonica DM, et al. Staging of primary colorectal carcinomas with fluorine-18 fluorodeoxyglucose whole-body PET: correlation with histopathologic and CT findings. *Radiology* 1998, **206**, 755–760.
- Eigtved A, Andersson AP, Dahlstrom K, et al. Use of fluorine-18 fluorodeoxyglucose positron emission tomography in the detection of silent metastases from malignant melanoma. *Eur J Nucl Med* 2000, **27**, 70–75.
- Acland KM, O'Doherty MJ, Russell-Jones R. The value of positron emission tomography scanning in the detection of sub-clinical metastatic melanoma. *J Am Acad Dermatol* 2000, **42**, 606–611.
- Kau RJ, Alexiou C, Laubenbacher C, Werner M, Schwaiger M, Arnold W. Lymph node detection of head and neck squamous cell carcinomas by positron emission tomography with fluorodeoxyglucose F 18 in a routine clinical setting. *Arch Otolaryngol Head Neck Surg* 1999, **125**, 1322–1328.
- Mattei R, Rubello D, Ferlin G, Bagatella F. [Positron emission tomography (PET) with 18-fluorodeoxyglucose (FDG) in the diagnosis and preoperative staging of head and neck tumors: a prospective study]. Ruolo diagnostico della tomografia ad emissione di positroni (PET) con 18-fluorodesossiglucosio (FDG) nella stadiazione preoperatoria dei tumori del capo-collo: studio prospettico. *Acta Otorhinolaryngol Ital* 1998, **18**, 387–391.
- Bares R, Dohmen BM, Cremerius U, Fass J, Teusch M, Bull U. [Results of positron emission tomography with fluorine-18 labeled fluorodeoxyglucose in differential diagnosis and staging of pancreatic carcinoma] Ergebnisse der Positronenemissionstomographie mit Fluor-18-markierter Fluorodesoxyglukose bei Differentialdiagnose und Staging des Pankreaskarzinoms. *Radiologie* 1996, **36**, 435–440.
- Zinzani PL, Magagnoli M, Chierichetti F, et al. The role of positron emission tomography (PET) in the management of lymphoma patients. *Ann Oncol* 1999, **10**, 1181–1184.
- McGuirt WF, Greven K, Williams D, et al. PET scanning in head and neck oncology: a review. *Head Neck* 1998, **20**, 208–215.
- Stokkel MP, de Klerk JH, Zelissen PM, Koppeschaar HP, van Rijk PP. Fluorine-18 fluorodeoxyglucose dual-head positron emission tomography in the detection of recurrent differentiated thyroid cancer: preliminary results. *Eur J Nucl Med* 1999, **26**, 1606–1609.
- Grunwald F, Kalicke T, Feine U, et al. Fluorine-18 fluorodeoxyglucose positron emission tomography in thyroid cancer: results of a multicentre study. *Eur J Nucl Med* 1999, **26**, 1547–1552.
- Franke C, Klapdor R, Meyerhoff K, Schauman M. 18-FDG positron emission tomography of the pancreas: diagnostic benefit in the follow-up of pancreatic carcinoma. *Anticancer Res* 1999, **19**, 2437–2442.
- Takeuchi O, Saito N, Koda K, Sarashina H, Nakajima N. Clinical assessment of positron emission tomography for the diagnosis of local recurrence in colorectal cancer. *Br J Surg* 1999, **86**, 932–937.
- Asensio C, Perez-Castejon MJ, Maldonado A, et al. [The role of PET-FDG in questionable diagnosis of relapse in the presence of radionecrosis of brain tumors] Papel de la PET-FDG ante la duda diagnostica de recidiva frente a radionecrosis en tumores cerebrales. *Rev Neurol* 1998, **27**, 447–452.
- Bader JB, Samnick S, Schaefer A, et al. [Contribution of nuclear medicine to the diagnosis of recurrent brain tumors and cerebral radionecrosis]. Beitrag der Nuklearmedizin zur Diagnostik des Hirntumorrezidivs und der zerebralen Radionekrose. *Radiologie* 1998, **38**, 924–929.
- Kitagawa Y, Sadato N, Azuma H, et al. FDG PET to evaluate combined intra-arterial chemotherapy and radiotherapy of head and neck neoplasms. *J Nucl Med* 1999, **40**, 1132–1137.
- Sakamoto H, Nakai Y, Ohashi Y, et al. Monitoring of response to radiotherapy with fluorine-18 deoxyglucose PET of head and neck squamous cell carcinomas. *Acta Otolaryngol Suppl* 1998, **538**, 254–260.

37. Mitsuhashi N, Hayakawa K, Hasegawa M, *et al.* Clinical FDG-PET in diagnosis and evaluation of radiation response of patients with nasopharyngeal tumor. *Anticancer Res* 1998, **18**, 2827–2832.
38. Brock CS, Young H, O'Reilly SM, *et al.* Early evaluation of tumour metabolic response using [¹⁸F]fluorodeoxyglucose and positron emission tomography: a pilot study following the phase II chemotherapy schedule for temozolomide in recurrent high-grade gliomas. *Br J Cancer* 2000, **82**, 608–615.
39. Schulte M, Brecht-Krauss D, Werner M, *et al.* Evaluation of neoadjuvant therapy response of osteogenic sarcoma using FDG PET. *J Nucl Med* 1999, **40**, 1637–1643.
40. Schelling M, Avril N, Nahrig J, *et al.* Positron emission tomography using [(18)F]fluorodeoxyglucose for monitoring primary chemotherapy in breast cancer. *J Clin Oncol* 2000, **18**, 1689–1695.
41. Smith IC, Welch AE, Hutcheon AW, *et al.* Positron emission tomography using [(18)F]-fluorodeoxy-D-glucose to predict the pathologic response of breast cancer to primary chemotherapy. *J Clin Oncol* 2000, **18**, 1676–1688.
42. Safa AA, Tran LM, Rege S, *et al.* The role of positron emission tomography in occult primary head and neck cancers. *Cancer J Sci Am* 1999, **5**, 214–218.
43. Mukherji SK, Drane WE, Mancuso AA, Parsons JT, Mendenhall WM, Stringer S. Occult primary tumors of the head and neck: detection with 2-[F-18] fluoro-2-deoxy-D-glucose SPECT. *Radiology* 1999, **199**, 761–766.
44. Goldman S, Levivier M, Pirotte B, *et al.* Regional methionine and glucose uptake in high-grade gliomas: a comparative study on PET-guided stereotactic biopsy. Published erratum appears in *J Nucl Med* 1997 Dec, 38(12), 2002. *J Nucl Med* 1997, **38**, 1459–1462.
45. Pirotte B, Goldman S, Bidaut LM, *et al.* Use of positron emission tomography (PET) in stereotactic conditions for brain biopsy. *Acta Neurochir (Wien)* 1995, **134**, 79–82.
46. Lyden D, Young AZ, Zagzag D, *et al.* Id1 and Id3 are required for neurogenesis, angiogenesis and vascularization of tumour xenografts. *Nature* 1999, **401**, 670–677.
47. Brun E, Ohlsson T, Erlandsson K, *et al.* Early prediction of treatment outcome in head and neck cancer with 2-18FDG PET. *Acta Oncol* 1997, **36**, 741–747.
48. Price P. Positron emission tomography (PET) in diagnostic oncology. Is it a necessary tool today? *Eur J Cancer* 2000, **36**, 691–693.
49. Frackowiak RS, Lenzi GL, Jones T, Heather JD. Quantitative measurement of regional cerebral blood flow and oxygen metabolism in man using 15O and positron emission tomography: theory, procedure, and normal values. *J Comput Assist Tomogr* 1980, **4**, 727–736.
50. Herscovitch P, Markham J, Raichle ME. Brain blood flow measured with intravenous H₂(15)O. I. Theory and error analysis. *J Nucl Med* 1983, **24**, 782–789.
51. Wilson CB, Lammertsma AA, McKenzie CG, Sikora K, Jones T. Measurements of blood flow and exchanging water space in breast tumors using positron emission tomography: a rapid and noninvasive dynamic method. *Cancer Res* 1992, **52**, 1592–1597.
52. Vander Borgh T, Lambotte LE, Pauwels SA, Dive CC. Uptake of thymidine labeled on carbon 2: a potential index of liver regeneration by positron emission tomography. *Hepatology* 1990, **12**, 113–118.
53. Vander Borgh T, Lambotte L, Pauwels S, Labar D, Becker G, Dive C. Noninvasive measurement of liver regeneration with positron emission tomography and [2-11C]thymidine. *Gastroenterology* 1991, **101**, 794–799.
54. Price P, Harte RJA, Tilsley O, *et al.* The use of radiolabelled anticancer drugs in phase I/II clinical trials and the assessment of therapeutic efficacy of new agents using PET. In Comar D, ed. *PET for Drug Development and Evaluation*. The Netherlands, Kluwer Academic Publishers, 1995, 301–326.
55. Shields AF, Grierson JR, Dohmen BM, *et al.* Imaging proliferation in vivo with [F-18]FLT and positron emission tomography. *Nature Med* 1998, **4**, 1334–1336.
56. Shields AF, Mankoff DA, Link JM, *et al.* Carbon-11-thymidine and FDG to measure therapy response. *J Nucl Med* 1998, **39**, 1757–1762.
57. Bergstrom M, Lu L, Fasth KJ, *et al.* In vitro and animal validation of bromine-76-bromodeoxyuridine as a proliferation marker. *J Nucl Med* 1998, **39**, 1273–1279.
58. Tewson TJ, Krohn KA. PET radiopharmaceuticals: state-of-the-art and future prospects. *Semin Nucl Med* 1998, **28**, 221–234.
59. Yasukawa T, Yoshikawa K, Aoyagi H, *et al.* Usefulness of PET with 11C-methionine for the detection of hilar and mediastinal lymph node metastasis in lung cancer. *J Nucl Med* 2000, **41**, 283–290.
60. Nuutinen J, Jyrkkio S, Lehtikainen P, Lindholm P, Minn H. Evaluation of early response to radiotherapy in head and neck cancer measured with [11C]methionine-positron emission tomography. *Radiother Oncol* 1999, **52**, 225–232.
61. Ishiwata K, Kubota K, Murakami M, *et al.* Re-evaluation of amino acid PET studies: can the protein synthesis rates in brain and tumor tissues be measured in vivo? *J Nucl Med* 1993, **34**, 1936–1943.
62. Dehdashti F, Mortimer JE, Siegel BA, *et al.* Positron tomographic assessment of estrogen receptors in breast cancer: comparison with FDG-PET and in vitro receptor assays. *J Nucl Med* 1995, **36**, 1766–1774.
63. Bonasera TA, O'Neil JP, Xu M, *et al.* Preclinical evaluation of fluorine-18-labeled androgen receptor ligands in baboons. *J Nucl Med* 1996, **37**, 1009–1015.
64. Silverman DH, Hoh CK, Seltzer MA, *et al.* Evaluating tumor biology and oncological disease with positron-emission tomography. *Semin Radiat Oncol* 1998, **8**, 183–196.
65. Yang DJ, Li C, Kuang LR, *et al.* Imaging, biodistribution and therapy potential of halogenated tamoxifen analogues. *Life Sci* 1994, **55**, 53–67.
66. Liu A, Carlson KE, Katzenellenbogen JA. Synthesis of high affinity fluorine-substituted ligands for the androgen receptor. Potential agents for imaging prostatic cancer by positron emission tomography. *J Med Chem* 1992, **35**, 2113–2129.
67. Hendrikse NH, de Vries EG, Eriks-Fluks L, *et al.* A new in vivo method to study P-glycoprotein transport in tumors and the blood-brain barrier. *Cancer Res* 1999, **59**, 2411–2416.
68. Hendrikse NH, Franssen EJ, van der Graaf WT, Vaalburg W, de Vries EG. Visualization of multidrug resistance in vivo. *Eur J Nucl Med* 1999, **26**, 283–293.
69. Levchenko A, Mehta BM, Lee JB, *et al.* Evaluation of 11C-colchicine for PET imaging of multiple drug resistance. *J Nucl Med* 2000, **41**, 493–501.
70. Brady F, Luthra S, Brown G, *et al.* Radiolabelled tracers and anticancer drugs for assessment of therapeutic efficacy using PET. *Curr Pharm Design* 2000, in press.
71. Rasey JS, Koh WJ, Evans ML, *et al.* Quantifying regional hypoxia in human tumors with positron emission tomography of [18F]fluoromisonidazole: a pretherapy study of 37 patients. *Int J Radiat Oncol Biol Phys* 1996, **36**, 417–428.
72. Casciari JJ, Graham MM, Rasey JS. A modeling approach for quantifying tumor hypoxia with [F-18]fluoromisonidazole PET time-activity data. *Med Phys* 1995, **22**, 1127–1139.
73. Koh WJ, Bergman KS, Rasey JS, *et al.* Evaluation of oxygenation status during fractionated radiotherapy in human nonsmall cell lung cancers using [F-18]fluoromisonidazole positron emission tomography. *Int J Radiat Oncol Biol Phys* 1995, **33**, 391–398.
74. Aboagye EO, Maxwell RJ, Kelson AB, *et al.* Preclinical evaluation of the fluorinated 2-nitroimidazole N-(2-hydroxy-3,3,3-trifluoropropyl)-2-(2-nitro-1-imidazolyl) acetamide (SR-4554) as a probe for the measurement of tumour hypoxia. *Cancer Res* 1997, **57**, 3314–3318.

75. Saleem A, Aboagye EO, Price P. *In vivo* monitoring of drugs using radiotracer techniques. *Adv Drug Delivery Rev* 2000, **41**, 21–39.
76. Moolten FL. Drug sensitivity (“suicide”) genes for selective cancer chemotherapy. *Cancer Gene Ther* 1994, **1**, 279–287.
77. Gambhir SS, Barrio JR, Herschman HR, Phelps ME. Assays for noninvasive imaging of reporter gene expression. *Nucl Med Biol* 1999, **26**, 481–490.
78. Alauddin MM, Shahinian A, Kundu RK, Gordon EM, Conti PS. Evaluation of 9-[(3-¹⁸F-fluoro-1-hydroxy-2-propoxy)methyl]guanine ([¹⁸F]-FHPG) *in vitro* and *in vivo* as a probe for PET imaging of gene incorporation and expression in tumors. *Nucl Med Biol* 1999, **26**, 371–376.
79. Blasberg RG, Tjuvajev JG. Herpes simplex virus thymidine kinase as a marker/reporter gene for PET imaging of gene therapy. *QJ Nucl Med* 1999, **43**, 163–169.
80. Front D, Israel O, Iosilevsky G, *et al.* SPECT quantitation of cobalt-57 bleomycin delivery to human brain tumors. *J Nucl Med* 1988, **29**, 187–194.
81. Ginos JZ, Cooper AJ, Dhawan V, *et al.* [¹³N]cisplatin PET to assess pharmacokinetics of intra-arterial versus intravenous chemotherapy for malignant brain tumors. *J Nucl Med* 1987, **28**, 1844–1852.
82. Mitsuki S, Diksic M, Conway T, Yamamoto YL, Villemure JG, Feindel W. Pharmacokinetics of ¹¹C-labelled BCNU and SarCNU in gliomas studied by PET. *J Neurooncol* 1991, **10**, 47–55.
83. Dimitrakopoulou A, Strauss LG, Clorius JH, *et al.* Studies with positron emission tomography after systemic administration of fluorine-18-uracil in patients with liver metastases from colorectal carcinoma. *J Nucl Med* 1993, **34**, 1075–1081.
84. Moehler M, Dimitrakopoulou-Strauss A, Gutzler F, Raeth U, Strauss LG, Stremmel W. ¹⁸F-labeled fluorouracil positron emission tomography and the prognoses of colorectal carcinoma patients with metastases to the liver treated with 5-fluorouracil. *Cancer* 1998, **83**, 245–253.
85. Dimitrakopoulou-Strauss A, Strauss LG, Schlag P, *et al.* Fluorine-18-fluorouracil to predict therapy response in liver metastases from colorectal carcinoma. *J Nucl Med* 1998, **39**, 1197–1202.
86. Harte RJ, Matthews JC, O’Reilly SM, *et al.* Tumor, normal tissue, and plasma pharmacokinetic studies of fluorouracil biomodulation with N-phosphonacetyl-L-aspartate, folinic acid, and interferon alfa. *J Clin Oncol* 1999, **17**, 1580–1588.
87. Saleem A, Yap J, Osman S, *et al.* Modulation of fluorouracil tissue pharmacokinetics by eniluracil: in-vivo imaging of drug action. *Lancet* 2000, **355**, 2125–2131.
88. Ghielmini M, Schmitz SF, Burki K, *et al.* The effect of Rituximab on patients with follicular and mantle-cell lymphoma. Swiss Group for Clinical Cancer Research (SAKK). *Ann Oncol* 2000, **11**(Suppl. 1), 123–126.
89. Cobleigh MA, Vogel CL, Tripathy D, *et al.* Multinational study of the efficacy and safety of humanized anti-HER2 monoclonal antibody in women who have HER2-overexpressing metastatic breast cancer that has progressed after chemotherapy for metastatic disease. *J Clin Oncol* 1999, **17**, 2639–2648.
90. Lubberink M, Lundqvist H, Westlin JE, *et al.* Positron emission tomography and radioimmunotargeting — aspects of quantification and dosimetry. *Acta Oncol* 1999, **38**, 343–349.